

Optimising Cervical Cancer Screening: A Critical Review of Quality Control, Assurance, and Process Improvement Strategies in Pap Smear Cytology

GRACY JEBA RANI¹, S SOWMYA², ES KEERTHIKA SRI³

ABSTRACT

Cervical cancer is a significant global health concern, with the Pap smear serving as a crucial tool for early detection. However, the effectiveness of Pap smear screening is compromised by false positives and negatives, necessitating robust Quality Control (QC) and Quality Assurance (QA) measures. This review examines the preanalytical, analytical, and post-analytical phases of Pap smear cytology, focusing on quality indicators that enhance diagnostic accuracy. The 2014 refinement of the Bethesda System for Reporting Cervical Cytology (BSRCC) provides a standardised framework for classifying cytological abnormalities. Key quality indicators, such as the ASCUS/SIL ratio and Cytology-Histology Concordance (CHC), are highlighted, along with the importance of Six Sigma metrics and Lean principles for evaluating and improving Pap smear procedures. The review emphasises the implementation of QC, QA, Six Sigma, and Lean principles in healthcare systems to ensure high diagnostic standards, operational efficiency, and patient safety.

Keywords: Cytology, Quality assurance, Quality parameters

INTRODUCTION

Cervical cancer is the fourth most common cancer in women globally, accounting for approximately 350,000 deaths in 2022, with an estimated 660,000 new cases by March 2024 [1]. In India, it is a leading cause of death among women aged 30-69, representing approximately 17% of mortality in this age group. The high prevalence—approximately 1 in 53 women testing positive compared to 1 in 100 in more developed countries—highlights its societal impact. While most cases exhibit slow progression (10-12 years from mild dysplasia to carcinoma) [2], early diagnosis and treatment are crucial for reducing mortality. The Pap smear, introduced in 1945 [3], remains an effective screening tool, encompassing pre-analytical, analytical, and post-analytical phases, and reported using the Bethesda system. This system classifies samples as low-grade or high-grade carcinomas (e.g., Atypical Squamous Cells (ASC) of Undetermined Significance (ASC-US), ASC-H, Low-grade Squamous Intraepithelial Lesion (LSIL), High-grade Squamous Intraepithelial Lesion (HSIL), Atypical Glandular Cells (AGC), squamous cell carcinoma, or adenocarcinoma). Despite its costeffectiveness and contribution to reduced mortality, discrepancies in results lead to a high number of false positives, underscoring the need for QC and QA measures [4].

QC, initially developed in industry, has expanded to healthcare laboratories, evolving from clinical chemistry to other disciplines, including QA in clinical settings [5]. Gynaecological cytology is subject to stringent regulations, with QA enforced by the Clinical Laboratory Improvement Amendments of 1988 (CLIA '88). QA in Pap smear cytology aims to improve test performance, reducing false negatives (which can lead to advanced disease and mortality) and false positives [6]. Quality indicators monitor the QA system, transforming laboratory medicine. Internal QC detects non-conformities throughout the laboratory process [7]. These indicators include positivity rate, percentage of ASC-compatible tests, ASCUS/SIL ratio, percentage of LSIL/HSIL-compatible tests, false negative rates, and unsatisfactory smear percentages [4,8,9].

Quality indicators ensure accurate detection of precancerous lesions, preventing false positives and negatives. Pap smear audits systematically review and evaluate reports and processes to assess quality, accuracy, and effectiveness, identifying deficiencies and areas for improvement [10,11]. Lean Six Sigma, a design approach with specialised tools, enhances quality and safety outcomes in healthcare [12]. In Pap smear cytology, it eliminates inefficiencies, minimises variation, and ensures consistent quality, improving patient outcomes and resource utilisation. This review emphasises the importance of QC, QA, quality indicators, internal QC, and Lean Six Sigma in Pap smear cytology.

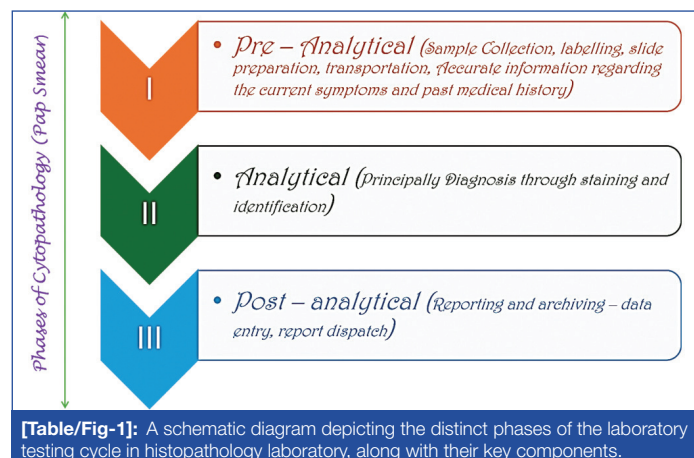
2. Critical Phases in Cytopathology for Cervical Cancer Diagnosis Using Pap Smears: Pre-Analytical, Analytical, and Post-Analytical Considerations

Pap smear processing involves three crucial phases: pre-analytical, analytical, and post-analytical [13]. The preanalytical phase, responsible for 49-68% of errors [14], involves specimen handling before it reaches the laboratory. This includes proper collection, fixation, and transport to preserve sample integrity. Stringent oversight, staff education, standardised procedures, QC measures, automation technologies (e.g., Hologic's ThinPrep Imaging System, Becton Dickinson's Focal Point GS Imaging System), and Computer-Aided Diagnosis (CAD) systems (e.g., Cytoc's AutoPap, Visioneer's Pathfinder) are crucial to reduce pre-analytical errors [14-18]. Rejecting unlabelled samples and documenting re-labelling with physician approval are also essential [14-18].

The analytical phase involves microscopic examination by cytotechnologists and pathologists to identify cellular morphology, including dysplastic cells, ASC, or cancerous lesions. Staining, identifying abnormalities, and diagnosing based on morphological criteria are key steps. Advanced techniques, such as computer-assisted screening and molecular diagnostics, enhance diagnostic accuracy [14-18].

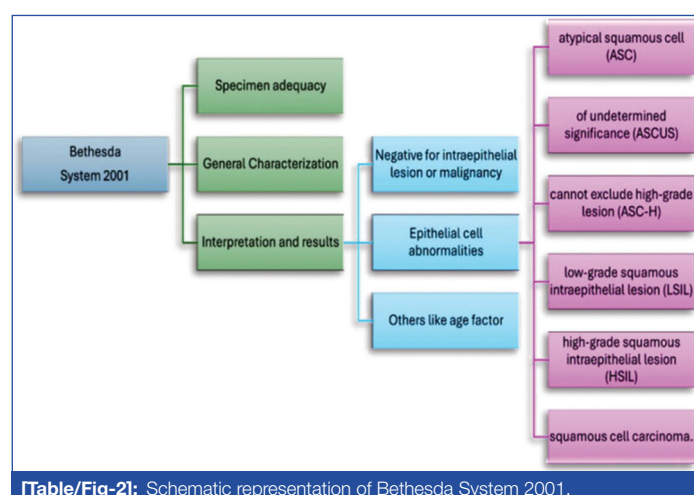
The post-analytical phase includes reporting results to clinicians, interpreting findings within the patient's clinical history, and ensuring appropriate follow-up. Effective communication and documentation

are vital for timely and appropriate patient care [19]. Reducing errors and enhancing quality requires analysis of analytical and postanalytical phases, continuous cyto-histopathological correlation, internal and external quality programs, timely turnaround with proper documentation, and continuous training [19]. Advanced Electronic Medical Record (EMR) systems (e.g., Epic EMR) facilitate electronic transmission of results, reducing transcription errors and Turnaround Time (TAT) [Table/Fig-1] [20,21]. Effective QC and QA are integral to all phases, ensuring high standards from sample collection to result reporting. The Food and Drug Administration (FDA) requires rescreeing of 10% of negative Pap smears, along with smears from high-risk patients [22]. QA focuses on the outcome of the entire process, relying on comprehensive quality management, including system design, policies, and processes to prevent false results and ensure patient safety.



3. Bethesda System for Reporting Cervical Cytology (BSRCC)

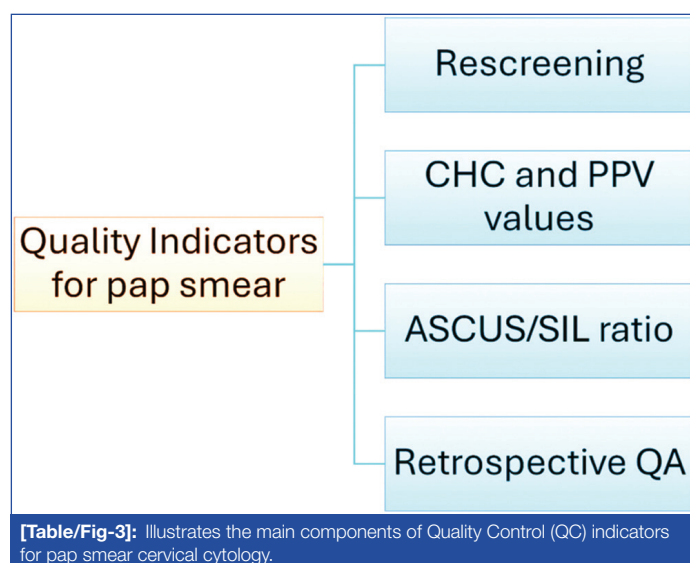
The BSRCC, developed in 1988, standardises reporting of cervical and vaginal cytology, classifying squamous cell abnormalities and providing clear criteria for specimen adequacy and cellular abnormality categorisation [4,9,23]. It supports timely identification of precancerous lesions [Table/Fig-2]. The system's primary purpose is to enhance clarity, consistency, and clinical utility of Pap smear interpretations [4,9,23]. By clearly defining categories (ASCUS, LSIL, HSIL), it enhances diagnostic precision, distinguishing between different degrees of cellular abnormalities [24]. The system also describes negative findings, including infections (*Trichomonas vaginalis*, *Candida* sp., *Actinomyces*, Herpes Simplex Virus (HSV)) and non-neoplastic findings (inflammation, atrophy) [24]. The BSRCC provides explicit recommendations for follow-up and treatment, aiding clinical decision-making [25]. The 2014 update incorporated new scientific insights and technological advancements, reflecting evolving understanding of cervical pathology [26]. The BSRCC enhances training and proficiency of cytopathologists and contributes to QA [27]. It distinguishes abnormalities due to HPV infection or condyloma



and various grades of dysplasia, and is well-positioned to incorporate future advancements [27].

4. The distinct phases of Pap smear cytology, guided by the standardised Bethesda System, ensure accurate diagnoses and effective communication, while quality indicators play a crucial role in maintaining procedural integrity.

Clinical laboratories must deliver precise and timely results. Cytological analysis is an essential tool for clinicians, aiding in treatment initiation and disease classification [28]. Total quality management is crucial for upholding quality standards, minimising errors, and enhancing patient safety [29]. The pre-analytical, analytical, and post-analytical processes are essential for the reliability and accuracy of Pap smears [30]. Evaluation of these phases reveals key insights into process quality and efficiency [5]. Quality indicators reflect the standards and effectiveness of patient care, serving as benchmarks for evaluating healthcare services [12,31]. They identify areas for improvement, maintain compliance, and enhance patient outcomes. Quality indicators for Pap smear cytology include rescreeing, ASCUS/Squamous Intraepithelial Lesion (SIL) ratio, retrospective QA, Positive Predictive Value (PPV), and CHC [Table/Fig-3]. Rescreeing requires random rescreeing of at least 10% of negative Pap smears, especially for high-risk patients [22]. The ASCUS/SIL ratio (number of ASCUS diagnoses/number of SIL diagnoses) is calculated, with a low ratio suggesting a more conservative approach to categorising ambiguous cases [7,8,32]. Retrospective QA involves reviewing past test results and procedures to identify errors and areas for improvement [13]. PPV (true positives/(true positives+false positives)×100%) indicates test accuracy [33]. CHC describes the agreement between cytological and histological findings [33]. A recent study highlights the importance of incorporating CHC and PPV as key quality indicators [34].

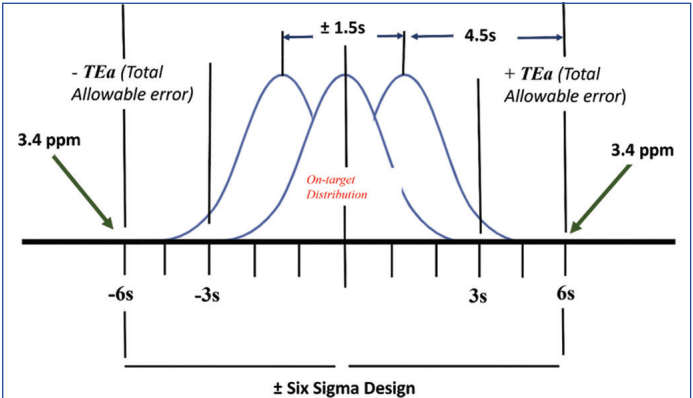


5. Understanding Non-Conformity in Healthcare: Implications for Quality Management and Patient Safety

Non-conformity or Non-Conforming Event (NCE) refers to deviations from established standards, guidelines, or expectations. These deviations can occur at various levels, including clinical processes, protocols, administrative procedures, and documentation [35]. Non-conformities encompass medication errors, diagnostic inaccuracies, patient safety incidents, regulatory violations, and failure to adhere to best practices [35,36]. Identifying and addressing non-conformities is crucial for patient safety, quality of care, and stakeholder trust [36]. The literature emphasises proactive risk management, quality improvement initiatives, and regulatory compliance to mitigate non-conformities [35-39]. CLIA '88 mandates rescreeing of at least 10% of negative cases (random and highrisk), retrospective review of HSIL or higher diagnoses over five years, and CHC [22].

6. Six Sigma metrics in clinical cytopathology enhance diagnostic accuracy by systematically reducing errors across pre-analytical, analytical, and post-analytical phases.

Six Sigma is a quality management methodology aimed at minimising defects and errors in laboratory processes. Developed in manufacturing, it has been adapted to healthcare, including clinical pathology, to enhance efficiency, accuracy, and patient safety [40-44]. Six Sigma principles provide a systematic approach for devising an effective QC strategy. Nevalainen et al., and Westgard conducted pioneering studies on evaluating laboratory performance using the Six Sigma scale [40,41]. The Six Sigma metric system (0-6) has a minimum acceptable quality level of 3 (66,807 faults per million), with 6 representing only 3.4 faults per million (99.99% success rate) [42]. A 1.5 standard deviation shift in the mean indicates a deviation from expected performance, potentially leading to increased variability and errors [Table/Fig-4] [43]. Sigma values are applicable to both qualitative and quantitative assays, providing standardised benchmarks [44]. Six Sigma uses a data-driven approach and statistical tools to identify and eliminate sources of variation, aiming for near-perfect performance. By setting rigorous quality standards and continuously monitoring and improving processes, Six Sigma helps clinical pathology laboratories deliver high-quality results, reduce turnaround times, and optimise resource utilisation, contributing to better patient outcomes and healthcare system efficiency [13]. Adherence to Six Sigma principles enables improvements in workflow, efficiency, accuracy, and the integration of technologies like telecytology and automation [13].



[Table/Fig-4]: This figure visually represents the quality of a Six Sigma assay, a standard measurement system used to assess process performance [41].

7. Integrating lean principles in hospital settings, focusing on cervical cytology to improve process efficiency, reduce errors, and ensure stringent quality

Lean principles, originating from lean manufacturing, aim to improve efficiency, quality, and patient outcomes in healthcare [45]. Lean focuses on eliminating waste, optimising processes, and enhancing value for patients. It emphasises continuous improvement and frontline staff involvement in problem-solving [46]. Lean strategies include streamlining workflows, reducing wait times, minimising unnecessary steps, and standardising procedures [47]. By eliminating inefficiencies, healthcare organisations improve patient satisfaction, reduce medical errors, and enhance quality of care [47,48]. Lean encourages innovation and collaboration [43]. Michael et al., created a Value Stream Map (VSM) of the Papanicolaou test procedure to identify areas for waste and error reduction. A redesigned VSM with Lean tools (first-in, first-out processes, minimised batch sizes) and staff engagement resulted in enhanced quality, increased patient safety, and greater operational efficiency [49]. Rabb et al., study showed that Lean workflows significantly reduced the proportion of Pap tests lacking a transformation zone component and increased the detection of cervical intraepithelial neoplasia, without significantly changing unsatisfactory Pap tests or ASC of unknown significance [50].

8. Enhancing Laboratory Performance in Pap Smear Cytology: Mitigating Non-Conformities through Rigorous Quality Control (QC) and Advanced Training Based on Comprehensive Literature Review.

A literature review on internal QC indicators for cervical smears revealed a limited number of comprehensive studies [Table/Fig-5] [7,19,51-55]. Most non-conformities occur during the pre-analytical phase, with errors such as inadequate or incorrect clinical information on test request forms having a substantial impact [7,19]. Inadequate technical training of personnel contributes to these non-conformities. Addressing these challenges requires comprehensive technical training programs focusing on proper specimen handling, accurate documentation, and effective communication. Ensuring detailed clinical information (patient demographics, medical history, reasons for testing) is critical for precise cytological interpretation. Institutions must enforce stringent protocols for thorough and complete data entry (preferably automated) before processing Pap smears [7,19]. CYTOTRAIN, a European Commission-funded program, trained doctors, cytopathologists, and cytotechnicians to maintain high-quality standards and minimise false positives and negatives. However, regular updates and training are necessary to keep professionals up-to-date [7,19].

S. No.	Pre-analytical phase	Analytical phase	Post analytical phase	Duration of study	Authors
1	57%	11%	32%	5 years	Chandra S et al., 2019 [19]
2	62.1%	NA	The study was not referenced in this phase	1 year	Muthukrishnan R et al., 2020 [51]
3	The study was not referenced in this phase	3.34%	The study was not referenced in this phase	3 months	Rajagopal P et al., 2023 [7]
4	4.5%	The study was not referenced in this phase	The study was not referenced in this phase	2 years	Belekar SV et al., 2023 [52]
5	The study was not referenced in this phase	Discrepancy in interobservation time	The study was not referenced in this phase	3 months	Siddegowda RB et al., 2016 [53]
6	Lack of pertinent clinical information on Pap test request form	The study was not referenced in this phase	The study was not referenced in this phase	Comprehensive literature synthesis	Kumar N et al., 2020 [54]
7	24.42%	The study was not referenced in this phase	The study was not referenced in this phase	1 year	Narasimha A et al., 2011 [55]

[Table/Fig-5]: Comparison of internal Quality Control (QC) indicators for cervical smears in various studies [7,19,51-55].

CONCLUSION(S)

Standardised protocols and continuous training minimise variability and enhance the quality of smear preparation, fixation, and staining. The integration of QC measures, including Six Sigma metrics and Lean principles, significantly enhances diagnostic accuracy, patient safety, and operational efficiency. Focusing on critical pre-analytical and analytical phases and utilising frameworks like the BSRCC streamlines processes and reduces non-conformity. These advancements ensure that Pap smear procedures remain reliable and precise, reinforcing their role in early detection and prevention of cervical cancer.

REFERENCES

- [1] Pimple S, Mishra G. Cancer cervix: Epidemiology and disease burden. *Cytojournal*. 2022;19:21.
- [2] Schiffman M, Castle PE, Jeronimo J, Rodriguez AC, Wacholder S. Human papillomavirus and cervical cancer. *Lancet*. 2007;370(9590):890-907.
- [3] Koss LG. Cervical (Pap) smear. New directions. *Cancer*. 1993;71(4):1406-12.
- [4] Davey DD, Nielsen ML, Naryshkin S, Robb JA, Cohen T, Kline TS. Atypical squamous cells of undetermined significance. Current laboratory practices of participants in the College of American Pathologists Interlaboratory Comparison program in cervicovaginal cytology. *Arch Pathol Lab Med*. 1996;120(5):440-44.
- [5] Branca M, Longatto-Filho A. Recommendations on quality control and quality assurance in cervical cytology. *Acta Cytol*. 2015;59(5):361-69.
- [6] Crothers BA, Booth CN, Darragh TM, Zhao C, Souers RJ, Thomas N, et al. False-positive Papanicolaou (PAP) test rates in the College of American Pathologists PAP education and PAP proficiency test programs: Evaluation of false-positive responses of high-grade squamous intraepithelial lesion or cancer to a negative reference diagnosis. *Arch Pathol Lab Med*. 2014;138:613-19.
- [7] Rajagopal P, Shankaralingappa A, Vijayan S. Internal quality control indicators in cervical smear screening- report from a tertiary care centre, India. *J Clin of Diagn Res*. 2023;17(4):EC01-EC04.
- [8] Araujo MLC, Santana DA, Almeida LB. Quality in cytopathology: An analysis of the internal quality monitoring indicators of the Instituto Nacional de Cancer. *J Bras Patol Med Lab*. 2015;51(2):102-07.
- [9] Kurman RJ, Solomon D. The Bethesda System for Reporting Cervical/Vaginal Cytologic Diagnoses. New York, NY: Springer Verlag; 1994:34.
- [10] Nayar R, David CW. The Bethesda System for Reporting Cervical Cytology. Springer, 2015.
- [11] American Society for Colposcopy and Cervical Pathology (ASCCP). "Quality Improvement Guidelines for Screening, Diagnosis, and Management of Cervical Intraepithelial Neoplasia (CIN)." ASCCP, 2019.
- [12] Pathology Outlines. Management Label Workflow. Available from: <https://www.pathologyoutlines.com/topic/managementlabelworkflow.html>. Accessed June 26, 2024.
- [13] Jum'ah H, Ganesan S. Quality assurance for cytopathology. *PathologyOutlines.com* Available from: <https://www.pathologyoutlines.com/topic/managementlabappdigpathqa.html>. Accessed June 4th, 2024.
- [14] Kani V, Kavitha K, Sonti S. Assessment of pre-analytical errors and fostering strategies to enhance accurate results and efficient turnaround times in the cytology laboratory of a tertiary care hospital. *Cureus*. 2024;16(3):e56592.
- [15] Bengtsson E, Malm P. Screening for cervical cancer using automated analysis of PAP-smears. *Comput Math Methods Med*. 2014;2014:842037.
- [16] Heckerman DE, Horvitz EJ, Nathwani BN. The pathfinder system. *Proc Annu Symp Comput Appl Med Care*. 1989;203-07.
- [17] Wilbur DC, Black-Schaffer WS, Luff RD, Abraham KP, Kemper C, Molina JT, et al. The Becton Dickinson FocalPoint GS Imaging System: Clinical trials demonstrate significantly improved sensitivity for the detection of important cervical lesions. *Am J Clin Pathol*. 2009;132(5):767-75.
- [18] Piaton E, Prat JJ, Nennig C, Hutin K, Colombel M, Ruffion A. ThinPrep® imaging system-assisted vs manual screening of urinary cytology slides in the detection of the "suspicious for high-grade urothelial carcinoma" category. *Cytopathology*. 2022;33(6):716-24.
- [19] Chandra S, Kusum A, Gaur DS, Chandra H. Analytical and post analytical phase of an ISO 15189:2012 certified cytopathology laboratory-a five year institutional experience. *J Cytol*. 2022;39(1):37-43.
- [20] Chishtie J, Sapiro N, Wiebe N, Rabatach L, Lorenzetti D, Leung AA, et al. Use of epic electronic health record system for healthcare research: Scoping review. *J Med Internet Res*. 2023;25:e51003.
- [21] Albagmi S. The effectiveness of EMR implementation regarding reducing documentation errors and waiting time for patients in outpatient clinics: A systematic review. *F1000Res*. 2021;10:514.
- [22] Marshall CJ, Rowe L, Bentz JS. Improved quality-control detection of false-negative Pap smears using the Autopap 300 QC system. *Diagn Cytopathol*. 1999;20(3):170-74.
- [23] Nayar R, Wilbur DC. The Bethesda System for Reporting cervical cytology: A historical perspective. *Acta Cytol*. 2017;61(4-5):359-72.
- [24] Solomon D, Davey D, Kurman R, Moriarty A, O'Connor D, Prey M, et al.; Forum Group Members; Bethesda 2001 Workshop. The 2001 Bethesda System: Terminology for reporting results of cervical cytology. *JAMA*. 2002;287(16):2114-19.
- [25] Pangarkar MA. The Bethesda System for reporting cervical cytology. *Cytojournal*. 2022;19:28.
- [26] Wang T, Zhang H, Liu Y, Zhao C. Updates in Cervical Cancer Screening Guidelines, The Bethesda System for Reporting Cervical Cytology, and Clinical Management Recommendations. *J Clin Transl Pathol*. 2023;3(2):75-83.
- [27] Nayar R, Wilbur DC. The Pap test and Bethesda 2014. *Cancer Cytopathol*. 2015;123(5):271-81.
- [28] de Moraes LSF, Magalhães JC, Braga IDS, Marega LA, Tavares SBDN, Amaral RG. Performance of laboratories after 10 years of participating in external quality monitoring in cervical cytology. *Acta Cytol*. 2020;64(3):224-31.
- [29] Ravikumar G. The need for quality control in cervical pap smear reporting. *J Interdiscip Histopathol*. 2022;10(7):01-02.
- [30] Chandra S, Chandra H, Kusum A, Singh Gaur D. Study of the pre-analytical phase of an ISO 15189: 2012-certified cytopathology laboratory: A 5-year institutional experience. *Acta Cytol*. 2019;63(1):56-62.
- [31] Wollersheim H, Hermens R, Hulscher M, Braspenning J, Ouwens M, Schouten J, et al. Clinical indicators: Development and applications. *Neth J Med*. 2007;65(1):15-22.
- [32] Renshaw AA, Deschênes M, Auger M. ASC/SIL ratio for cytotechnologists: A surrogate marker of screening sensitivity. *Am J Clin Pathol*. 2009;131(6):776-81.
- [33] Asaturova A, Dobrovolskaya D, Magnaeva A, Tregubova A, Bayramova G, Sukhikh G. Cervical Cytology-Histology Correlation Based on the American Society of Cytopathology Guideline (2017) at the Russian National Medical Research Center for Obstetrics, Gynecology, and Perinatology. *Diagnostics (Basel)*. 2022;12(1):210.
- [34] Senthil Kumar D, Ravikumar G. Quality metrics in cervical cytopathology: A single institutional study. *Cytopathology*. 2022;33(2):230-35.
- [35] Ho B, Ho E. The most common nonconformities encountered during the assessments of medical laboratories in Hong Kong using ISO 15189 as accreditation criteria. *Biochem Med (Zagreb)*. 2012;22(2):247-57.
- [36] Bruna L. Non-conformities that hinder health institution to be accredited: Integrative review. *Am J Biomed Res*. 2020;7:428-33.
- [37] Van Baarle E, Hartman L, Rooijackers S, Wallenburg I, Weenink JW, Bal R, et al. Fostering a just culture in healthcare organisations: Experiences in practice. *BMC Health Serv Res*. 2022;22(1):1035.
- [38] Pronovost PJ, Austin JM, Cassel CK, Delbanco SF, Jha AK, Kocher B, et al. Fostering transparency in outcomes, quality, safety, and costs. *Vital Directions for Health & Health Care: An Initiative of the National Academy of Medicine*. Washington (DC): National Academies Press (US); 2017.
- [39] Lee NY. Reduction of pre-analytical errors in the clinical laboratory at the University Hospital of Korea through quality improvement activities. *Clin Biochem*. 2019;70:24-29.
- [40] Nevalainen D, Berte L, Kraft C, Leigh E, Picaso L, Morgan T. Evaluating laboratory performance on quality indicators with the six sigma scale. *Arch Pathol Lab Med*. 2000;124(4):516-19.
- [41] Westgard JO. Six Sigma Quality Design and Control. Washington, DC: AACC Press; 2001.
- [42] Das S, Chaudhary N, Raju K. Analysis of the Six sigma principle in pre-analytical quality for hematological specimens. *Cureus*. 2023;15(7):e42434.
- [43] Shah Goel S, Saini R, Singh SB, Aggarwal O, Goel AK. Six sigma metrics and quality control in clinical laboratory. *Int J Med Res Rev*. 2014;2(2):140-49.
- [44] Badrick T. Sigma metrics. *Pathology*. 2017;49(Suppl 1):S15.
- [45] Lawal AK, Rotter T, Kinsman L, Sari N, Harrison L, Jeffery C, et al. Lean management in healthcare: Definition, concepts, methodology and effects reported (systematic review protocol). *Syst Rev*. 2014;3:103.
- [46] Bharsakade RS, Acharya P, Ganapathy L, Tiwari MK. A lean approach to healthcare management using multi criteria decision making. *OPSEARCH*. 2021;58(3):610-35.
- [47] Souza DL, Korzenowski AL, Alvarado MM, Sperafico JH, Ackermann AE, Mareth T, et al. A systematic review on lean applications' in emergency departments. *Healthcare (Basel)*. 2021;9(6):763.
- [48] Aij KH, Teunissen M. Lean leadership attributes: A systematic review of the literature. *J Health Organ Manag*. 2017;31(7-8):713-29.
- [49] Michael CW, Naik K, McVicker M. Value stream mapping of the Pap test processing procedure: A lean approach to improve quality and efficiency. *Am J Clin Pathol*. 2013;139(5):574-83.
- [50] Raab SS, Andrew-Jaja C, Grzybicki DM, Vrbin CM, Chesin CM, Fisch JM, et al. Dissemination of Lean methods to improve Pap testing quality and patient safety. *J Low Genit Tract Dis*. 2008;12(2):103-10.
- [51] Muthukrishnan R, Harshavardini S, Karthikeshwaran TM, Indumathy M. Evaluation of pre-analytical errors in cytology laboratory: A one-year study at a tertiary care hospital. *Int J Sci Res*. 2020;9(6):03-04.
- [52] Belekar SV, Kamal M, Warke AS. Cervical cytology and histology correlation as an analytic quality assurance exercise: Experience from an accredited cytology laboratory. *J Cytol*. 2023;40(4):205-10.
- [53] Siddegowda RB, DivyaRani MN, Natarajan M, Biligi DS. Inter-observer variation in reporting of pap smears. *National J Laboratory Med*. 2016;5(3):22-25.
- [54] Kumar N, Gupta R, Gupta S. Inadequate clinical data on Pap test request form: Where are we headed in the era of precision medicine? *CytoJournal*. 2020;17:1.
- [55] Narasimha A, Vasavi B, Harendra Kumar, Sapna M. An audit of pap smear cytology. *J South Asian Federat Obstetr Gynaecol*. 2011;3(3):121-24.

PARTICULARS OF CONTRIBUTORS:

- 1. Postgraduate, Department of Pathology, Sri Balaji Vidyapeeth, Pondicherry, India.
- 2. Professor and Head, Department of Pathology, Sri Balaji Vidyapeeth, Pondicherry, India.
- 3. Assistant Professor, Department of Pathology, Sri Balaji Vidyapeeth, Pondicherry, India.

NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

S Sowmya,
Mahatma Gandhi Medical College, Pillayarkuppam, Puducherry-607402, India.
E-mail: drssowmya@hotmail.com

PLAGIARISM CHECKING METHODS: [\[Jain H et al.\]](#)

- Plagiarism X-checker: Sep 12, 2024
- Manual Googling: Nov 12, 2024
- iThenticate Software: Jan 08, 2025 (10%)

ETYMOLOGY: Author Origin

EMENDATIONS: 7

AUTHOR DECLARATION:

- Financial or Other Competing Interests: None
- Was informed consent obtained from the subjects involved in the study? NA
- For any images presented appropriate consent has been obtained from the subjects. NA

Date of Submission: [Sep 09, 2024](#)

Date of Peer Review: [Oct 24, 2024](#)

Date of Acceptance: [Jan 10, 2025](#)

Date of Publishing: [May 01, 2025](#)